

# Histological Evaluation of Modern Cutting Instruments in Oral Mucosa

## A Comparative in Vitro Study

**Language:** English

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## Material and Methods

Porcine oral mucosa specimens were treated by a conventional scalpel, different laser wavelengths (Er:YAG, CO<sub>2</sub>, diode and Nd:YAG), high frequency surgery (hf1 SURG®, Vio® 300 D) and ultrasonic activated scalpels (UltraCision®, SonoSurg®) with evidence based settings. All specimens were fixed in 10% buffered formalin solution and embedded in paraffine. Sections were stained with hematoxylin-eosin (HE) and periodic acid-Schiff (PAS) reaction and examined light microscopically by two independent pathologists. The incisions were evaluated measuring the cutting efficacy, the width of damaged tissue adjacent to the incision and the quality of tissue and cells in the damaged areas. Finally the researched dates were described using synoptic gradings.

## Results

The cutting efficacy was demonstrated for all examined instruments. The width of the damage zones was within a range from the reference (scalpel = 0) to maximum values between 300 and 400 µm. In weaker magnification the damages were represented by the formation of hyalinized, eosinophilic zones along the defect borders and cleft or bubble formation in the peripheral epithelial and connective tissues. The histological changes were caused by mechanical and thermal influences. Altogether the conventional scalpel obtained best results, followed from laser and high frequency surgery with mid and ultrasonic scalpels with low acceptable results.

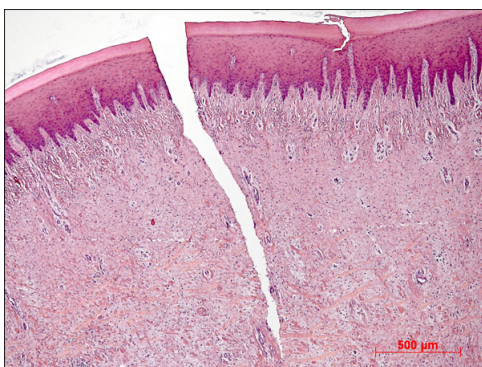


Fig. 1: Narrow, deep acute-angled defect (arrow) penetrating epithelium (e) and lamina propria (lp), smooth cutting margins, no peripheral tissue and cell damaging, H.E. staining

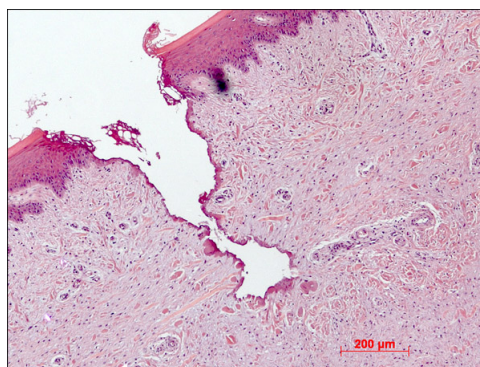


Fig. 2: Ampulla-like defect (arrow) penetrating epithelium (e) and lamina propria (lp); irregular cutting margins; narrow (5-40 µm) hyaline eosinophilic damage zone (asterisks), minimal peripheral tissue and cell damaging, H.E. staining

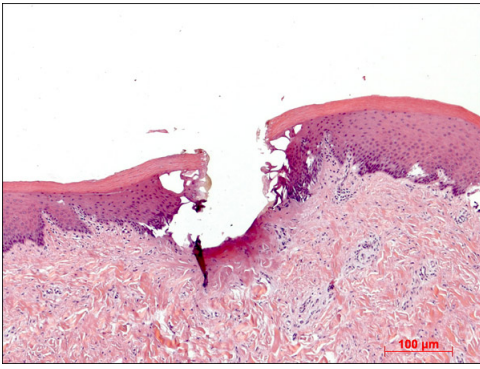


Fig. 3: Flat (100  $\mu$ m) defect (arrow) penetrating epithelium (e) and upper part of lamina propria (lp); hyaline eosinophilic damage zone (ca. 30  $\mu$ m, asterisks), minimal peripheral tissue and cell damaging, H.E. staining



Fig. 4: Deep defect (arrow) penetrating epithelium (e) and whole lamina propria (lp); irregular cutting margins; broad hyaline eosinophilic damage zone (300-400  $\mu$ m, asterisks), focal blistering (open arrow), H.E. staining

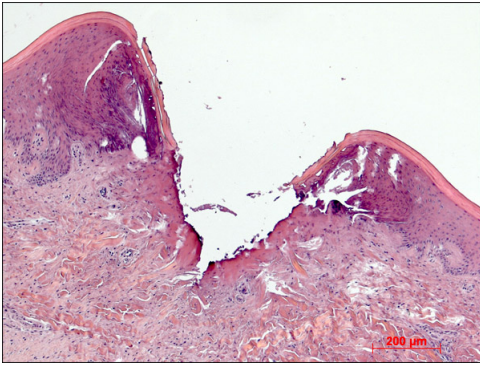


Fig. 5: Crater-like defect (arrow) penetrating epithelium (e) and lamina propria (lp); irregular cutting margins; hyaline eosinophilic damage zone (5-40  $\mu$ m, asterisks), intraepithelial clefts (open arrows), H.E. staining

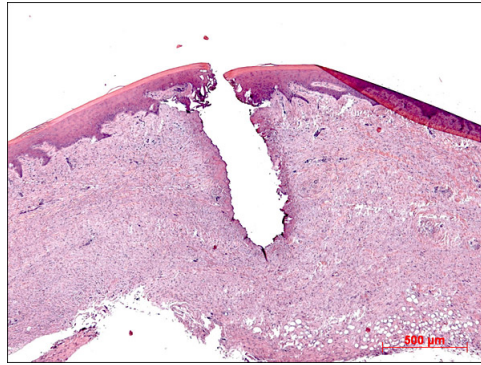


Fig. 6: Tube-like defect (arrow) penetrating epithelium (e) and deep into lamina propria (lp); irregular cutting margins; hyaline eosinophilic damage zone (50-100  $\mu$ m, asterisks), minimal peripheral tissue and cell damaging, H.E. staining

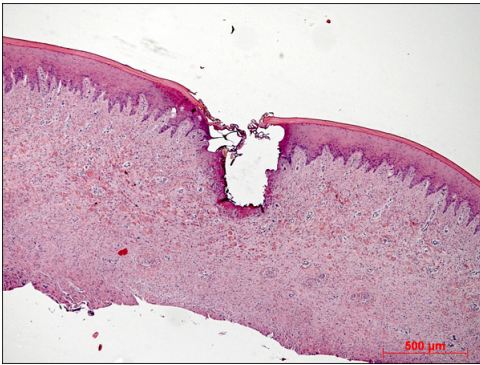


Fig. 7: Nearly rectangular defect (arrow) penetrating epithelium (e) and lamina propria (lp); irregular cutting margins; hyaline eosinophilic damage zone (10-50  $\mu$ m, asterisks), minimal peripheral tissue and cell damaging, carbonated epithelial rests projecting into cleft opening (open arrow); H.E. staining

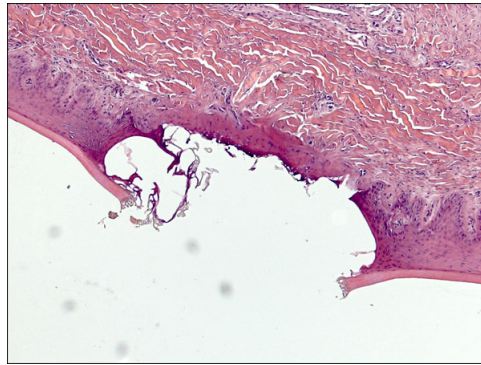


Fig. 8: Hollow defect (arrow) penetrating epithelium (e) and only apical parts of lamina propria (lp); irregular cutting margins; dense hyaline eosinophilic damage zone (ca. 50  $\mu$ m, asterisks), no peripheral tissue and cell damaging, carbonated epithelial rests projecting into cleft opening (open arrow); H.E. staining

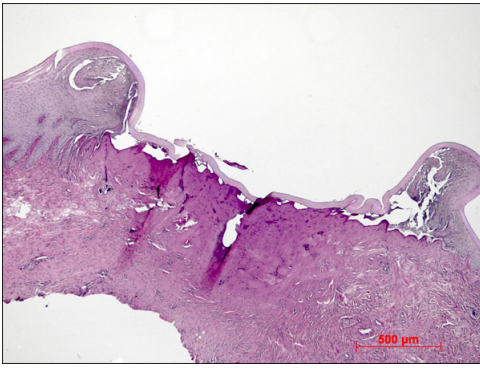


Fig. 9: Very hollow, massive defect (arrow) nearly only penetrating epithelium (e), which is rolled up at the defect borders), remnants of epithelial stratum corneum (c) covering the defect; dense hyaline PAS positive damage zone (ca. 50 µm, asterisks), massive peripheral tissue and cell damaging, e.g. cleft formations in epithelium (e) and lamina propria (lp); PAS staining

## Conclusions

The histological evaluation approved the application of established cutting instruments and the principal use of dissection techniques in oral mucosa. The potential advantages of interdisciplinary tools of soft tissue cutting instruments for oral surgery have yet to be better determined.

*This Poster was submitted by Prof. Dr. Dr. Winand Olivier.*

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# HISTOLOGICAL EVALUATION OF MODERN CUTTING INSTRUMENTS IN ORAL MUCOSA: A COMPARATIVE IN VITRO STUDY

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## BACKGROUND AND OBJECTIVE:

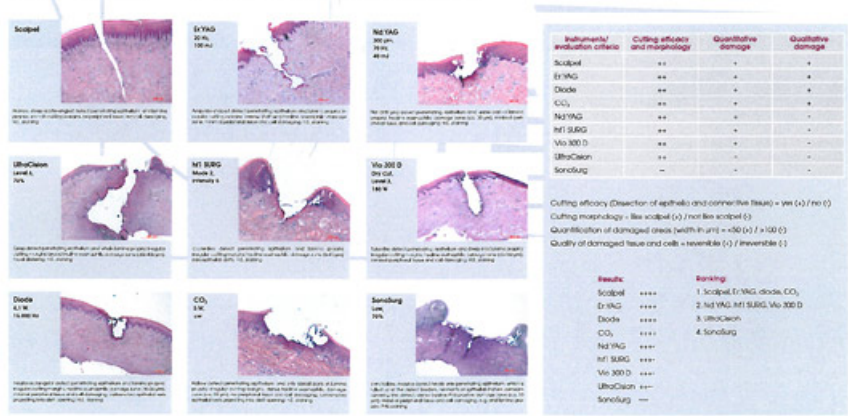
Cutting of soft tissues is an important procedure in oral surgery. Besides of established instruments exist various dissection techniques of minimally invasive surgery, which are not evaluated for oral applications. The aim of this in vitro experimental study was to perform a histological evaluation of incision effects produced in oral mucosa with modern cutting instruments of dental and visceral surgery.

## STUDY DESIGN:

Porcine oral mucosa specimens were treated by a conventional scalpel, different laser wavelengths (Er:YAG, CO<sub>2</sub> diode and Nd:YAG), high frequency surgery (HF SURG<sup>®</sup>, Vio<sup>®</sup> 300 D) and ultrasonic activated scalpels (UltraClison<sup>®</sup>, SonoSurg<sup>®</sup>) with evidence based settings. All specimens were fixed in 10% buffered formalin solution and embedded in paraffine. Sections were stained with hematoxylin-eosin (HE) and periodic acid-Schiff (PAS) reaction and examined light microscopically by two independent pathologists. The incisions were evaluated measuring the cutting efficacy, the width of damaged tissue adjacent to the incision and the quality of tissue and cells in the damaged areas. Finally the researched data were described using synoptic gradings.

## RESULTS:

The cutting efficacy was demonstrated for all examined instruments. The width of the damage zones was within a range from the reference (scalpel = 0) to maximum values between 300 and 400 µm. In softer magnification the damages were represented by the formation of hyalinized, eosinophilic zones along the defect borders and cleft or bubble formation in the peripheral epithelial and connective tissues. The histological changes were caused by mechanical and thermal influences. Altogether the conventional scalpel obtained best results, followed from laser and high frequency surgery with mid and ultrasonic scalpels with low acceptable results.



## CONCLUSIONS:

The histological evaluation approved the application of established cutting instruments and the principal use of dissection techniques in oral mucosa. The potential advantages of interdisciplinary tools of soft tissue cutting instruments for oral surgery have yet to be better determined.

**Keywords:**

Histological evaluation, oral mucosa, cutting instruments, in vitro study

**Senders' note:**

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